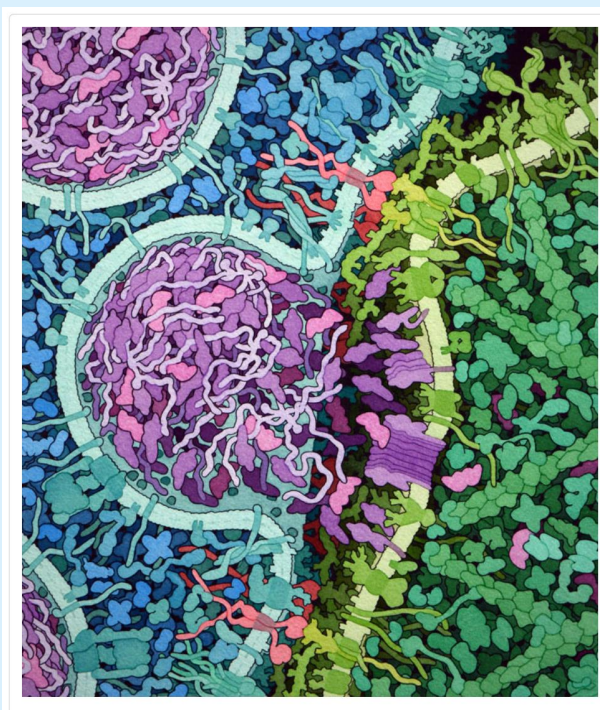
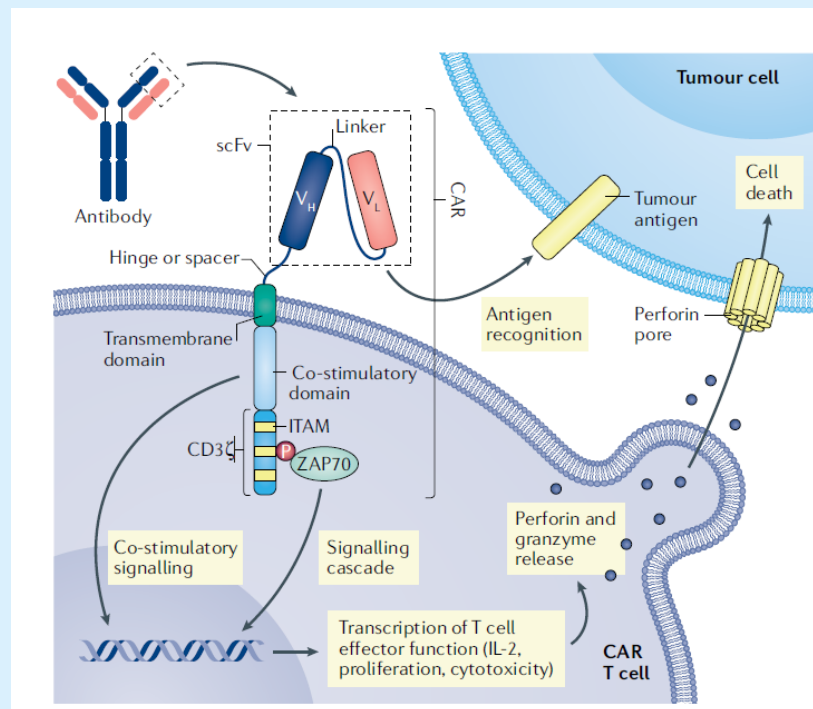


Living Drugs: *fighting cancer with CRISPR-engineered CAR-T cells.*



Workshop Outline

- **Cancer Immunotherapy**

Supercharging our own immune systems...

- **Connecting the dots.....**

- ***CRISPR-Based Genome Editing... just finished***

- ***Tomorrow's Science Today... coming soon***

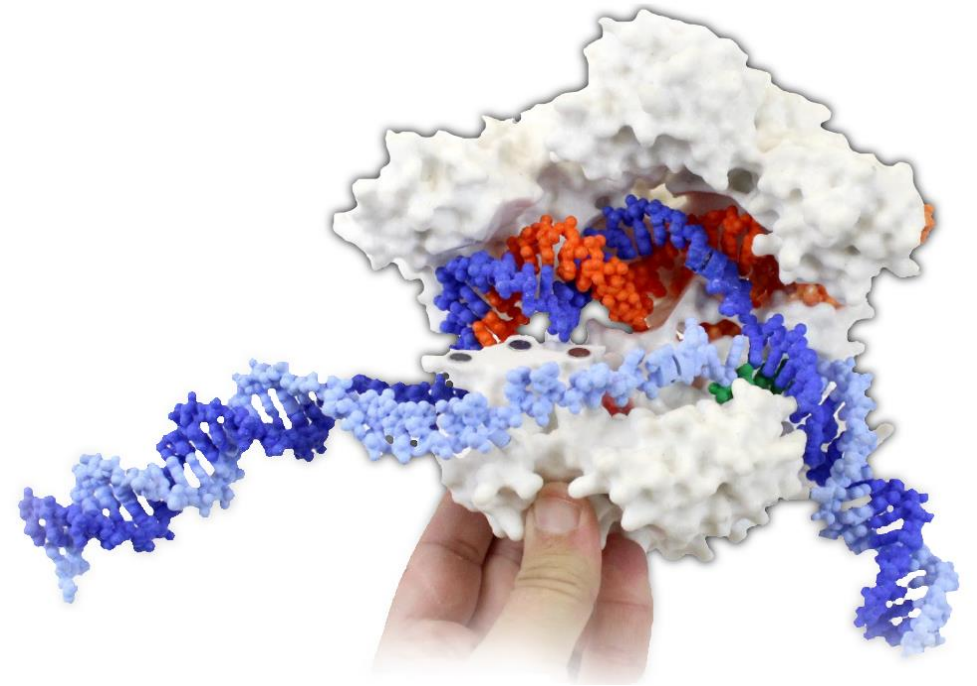
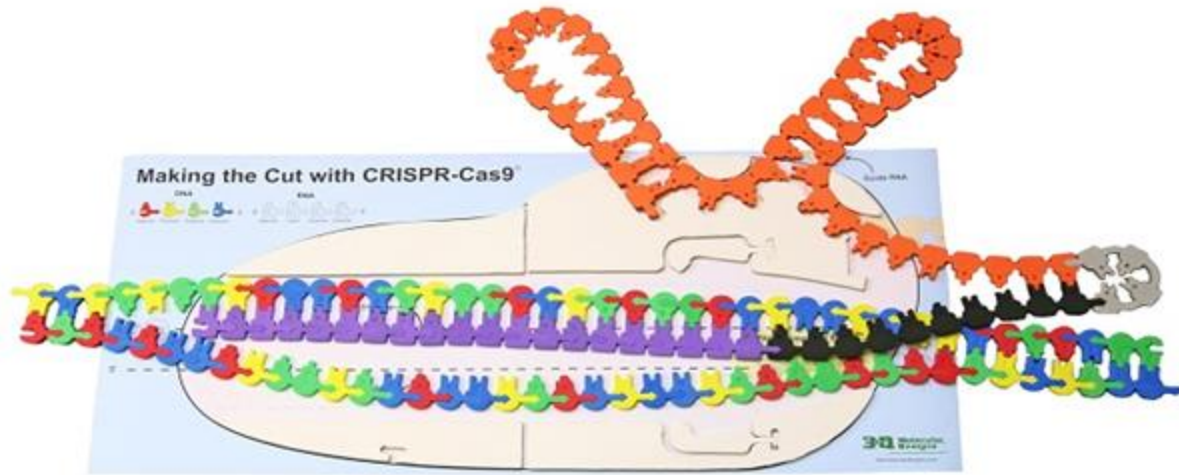
An NIH SEPA project -- focused on infectious diseases

- **What is a CAR-T Cell,... and how do we make them?**

Just completed....a 5-year NIH-SEPA project focused on:

The Science and Ethics of CRISPR-based Genome Editing

New tools that will be used to engineer proteins.... and cells.



CRISPR II: Using Cas9 as a Genome Editing Tool ---- at 1:20pm on Saturday

Coming soon,... to a workshop near you.

Tomorrow's Science Today: *preparing for the next pandemic.*

A new NIH SEPA project focused on infectious diseases

- **New Instructional Materials**

- Models, models and more models....
A Virus Collection,.. including CoV-2
Antibodies/vaccines /and **your immune system**
mRNA vaccines and multi-valent, ultra-potent vaccines...from synthetic biology.
 - Stories, stories and more stories....
- 
- ```
graph LR; A[Models, models and more models....
A Virus Collection,.. including CoV-2
Antibodies/vaccines /and your immune system
mRNA vaccines and multi-valent, ultra-potent vaccines...from synthetic biology.] --> B[T-Cells]; B --> C[CAR-T Cells]
```

- **A Professional Learning Experience**...beginning in the summer of 2024

But first,...enroll now in **Modeling the Molecular World, 2023**

Session 1 -- June 26–30, or

Session 2 -- July 17 – 21

# STEM....

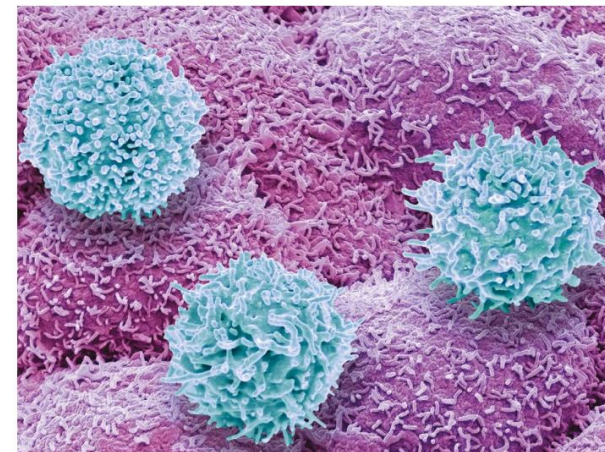
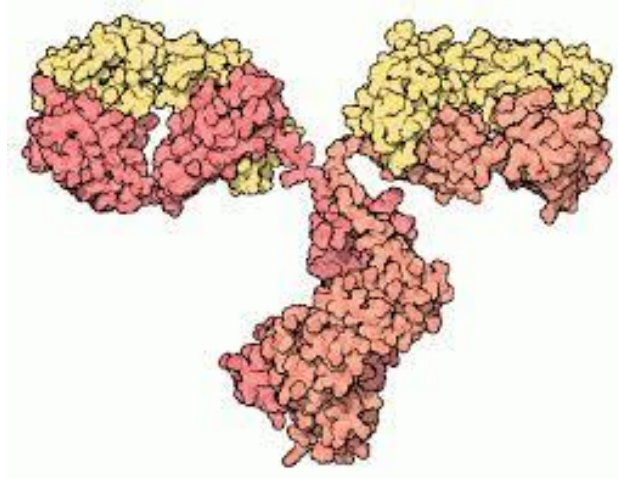
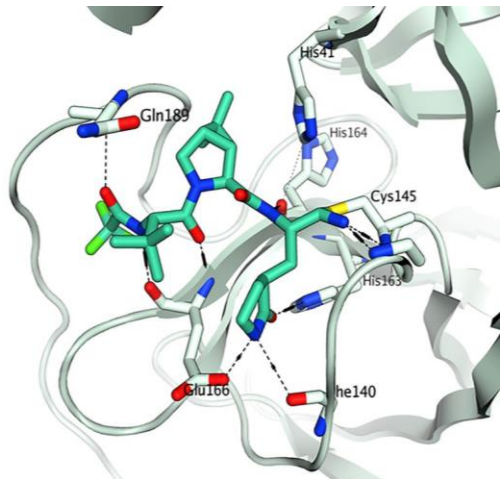
**E**ngineering... *allows us to:*

- \* *design an effective mRNA vaccine for CoV-2*
- \* *design supercharged T-cells to fight cancer*

**S**cience... *provided us with the nucleotide sequence of the CoV-2 spike protein.... and the field of molecular immunology*

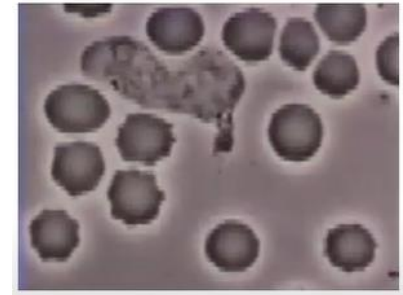
How can you incorporate principles of  
*engineering design* into your science classroom?

# Drugs,... Yesterday,... Today,... and Tomorrow



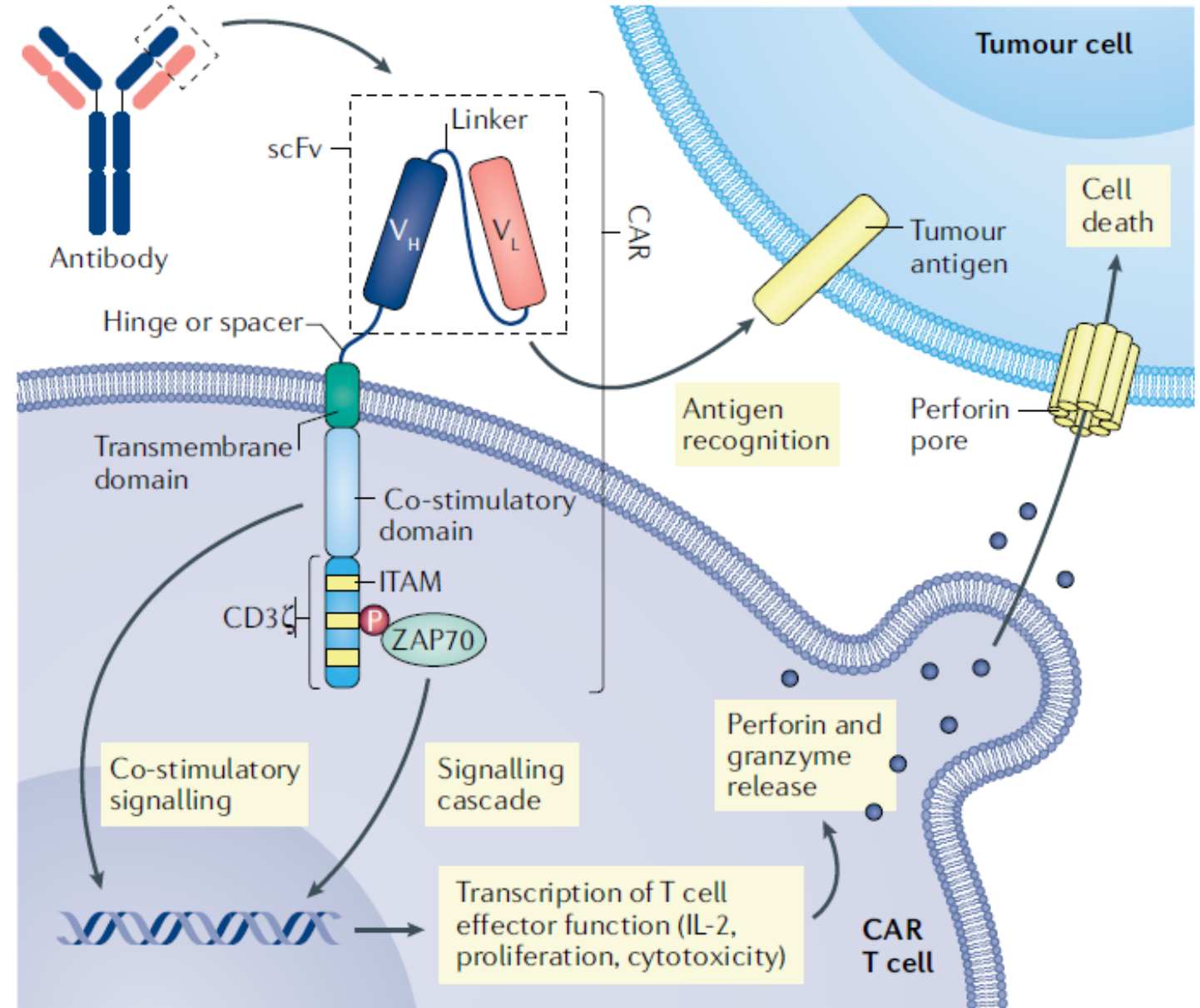
## Deciphering CAR-T Cells: Exploring Functional Mechanisms to Drive Next Generation Immunotherapy

A video...featuring Wendell Lim...  
<https://youtu.be/EAA-aSEZasw>  
start viewing at 24:20 ... or at 31:30

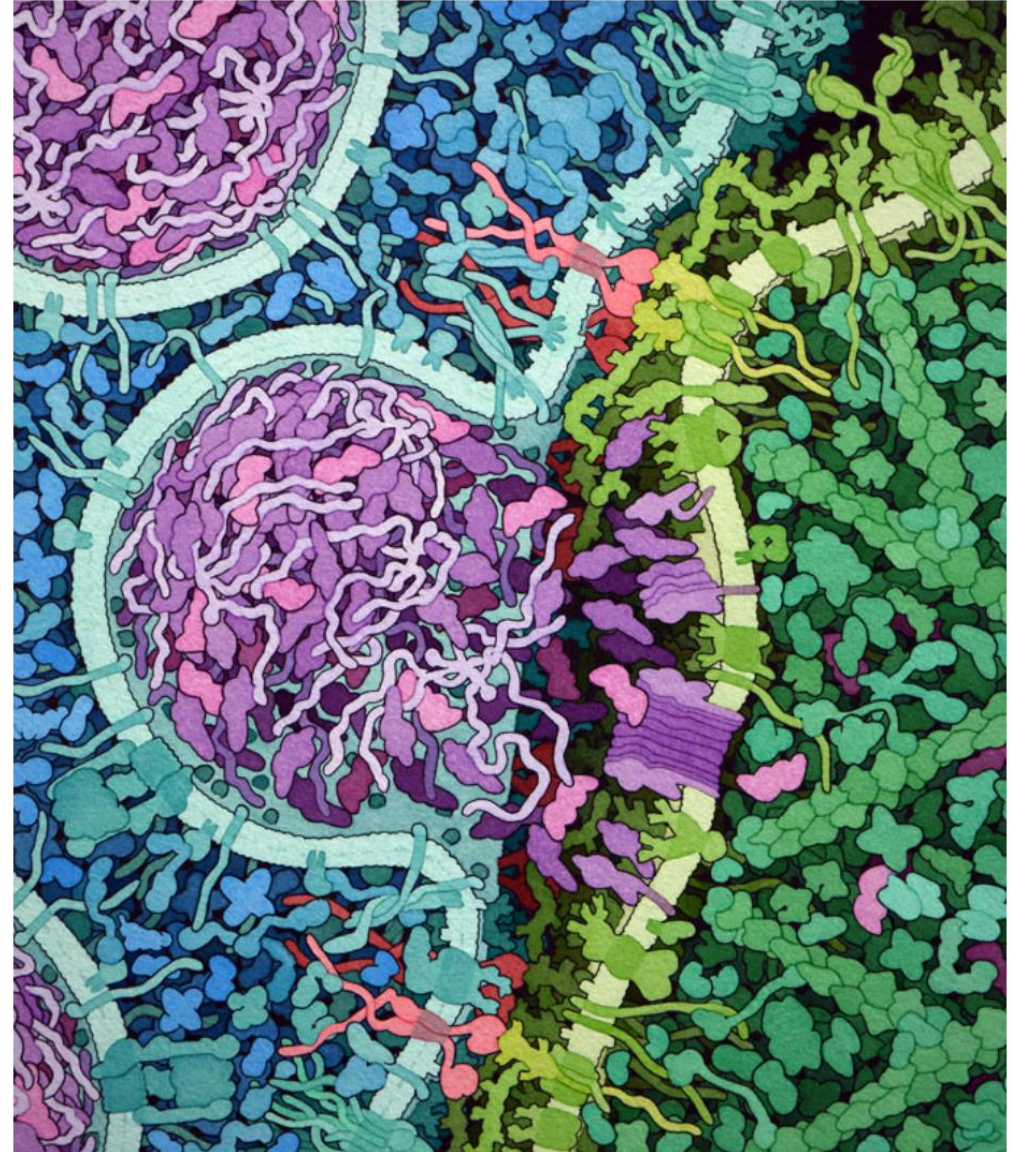


<https://www.youtube.com/watch?v=EAA-aSEZasw>

Fig. 1 | **Schematic of a basic second-generation CAR T cell.** The extracellular portion of the chimeric antigen receptor (CAR) molecule is typically generated from a monoclonal antibody against the target. The variable heavy ( $V_H$ ) and variable light ( $V_L$ ) chains, also known as the single-chain variable fragment (scFv), from the antibody sequence are connected by a linker to form the antigen-specific region of the CAR molecule. The hinge or spacer region anchors the scFv to the transmembrane region that traverses the cell membrane. Intracellularly, the co-stimulatory domain and CD3 $\zeta$  chain signal once the scFv portion of the CAR recognizes and binds tumour antigen. Co-stimulatory signals are dependent on the co-stimulation domain used: CD28 is dependent on PI3K, whereas 4-1BB requires tumour necrosis factor (TNF) receptor-associated factors (TRAFs) and nuclear factor- $\kappa$ B (NF- $\kappa$ B). The CD3 $\zeta$  chain contains three immunoreceptor tyrosine-based activation motif (ITAM) domains that, upon phosphorylation (P), signal through  $\zeta$ -associated protein of 70kDa (ZAP70). Downstream signalling leads to T cell effector functions including release of perforin and granzyme, leading to cell death of the target tumour cell. IL-2, interleukin 2.



*Artistic illustration of an engineered T cell (on the left side in blue) recognizing and attacking a leukemia cell (on the right side in green). The CAR molecule is shown in red, bound to CD19 on the leukemia cell. This has led to activation of the T cell, which releases perforin (purple), forming pores in the cell surface. Granzymes (magenta) then enter through the pore and initiate apoptosis to kill the cancer cell.*



## Molecule of the Month: Chimeric Antigen Receptors

*T cells may be engineered with chimeric antigen receptors to attack cancer cells.*

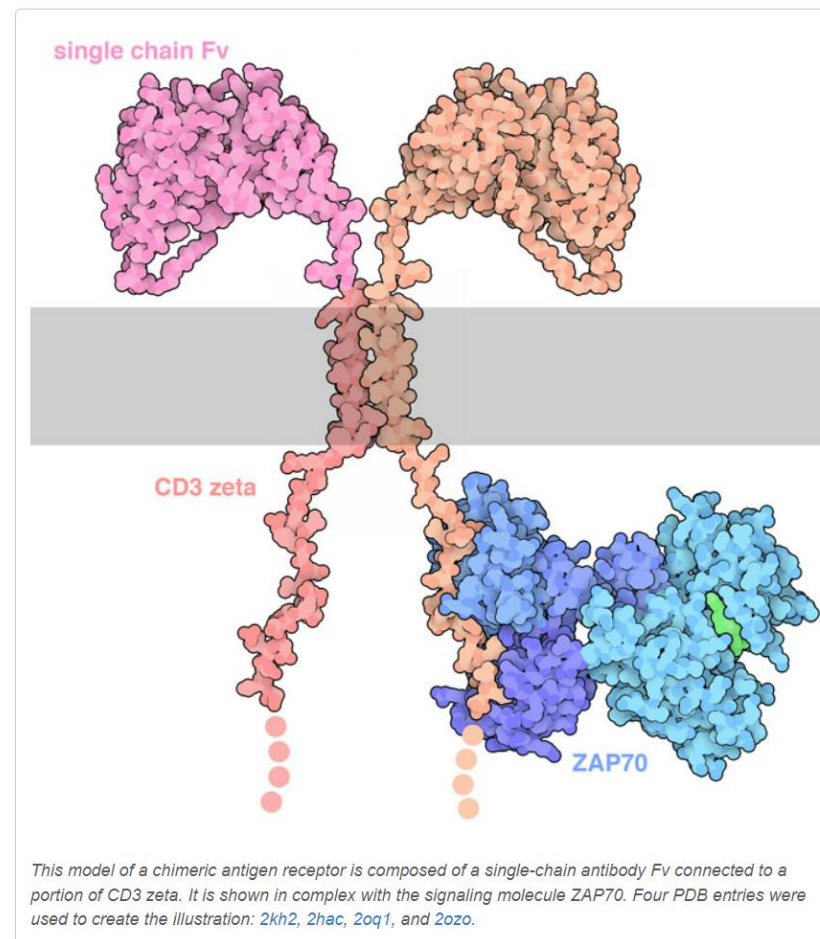
The immune system is expert at defending us from dangers, but when faced with cancer cells, the immune system often fails. Cancer cells are normal human cells that have been transformed into rogues. They usually have many genetic changes that allow them to grow without normal limitations. Unfortunately, most of these genetic changes mutate proteins that are inside the cancer cell, so the immune system may not detect them. Today, researchers are developing ways to retool the immune system to fight specific forms of cancer.

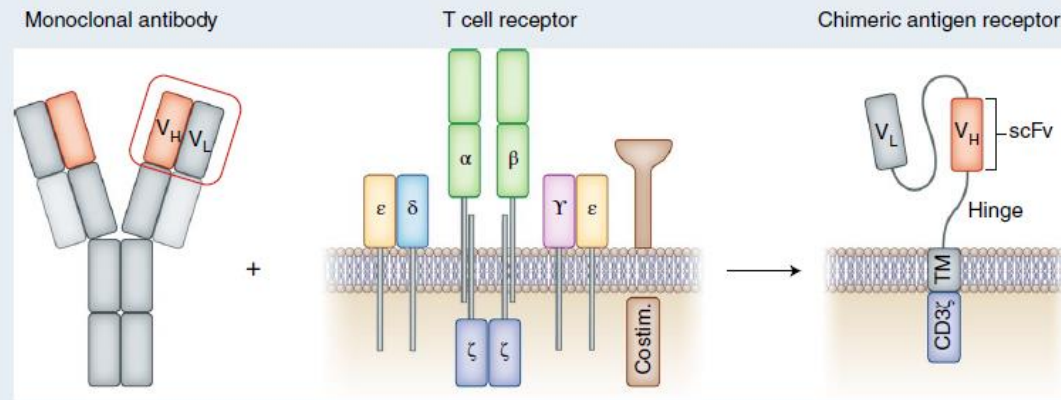
### Engineered T cells

T cells have the ability to recognize virus-infected cells using [T-cell receptors](#), and then destroy them. Unfortunately T cells are not as effective against cancer cells, because the molecules on the surface of cancer cells often look much like the molecules on normal cells. Chimeric antigen receptors (CAR) are a way for doctors to arm T cells with the ability to recognize cancer cells. Researchers remove T cells from a cancer patient and add a gene for a CAR that recognizes their form of cancer. When these “CART” cells are returned to the patient, they search for cancer cells and destroy them.

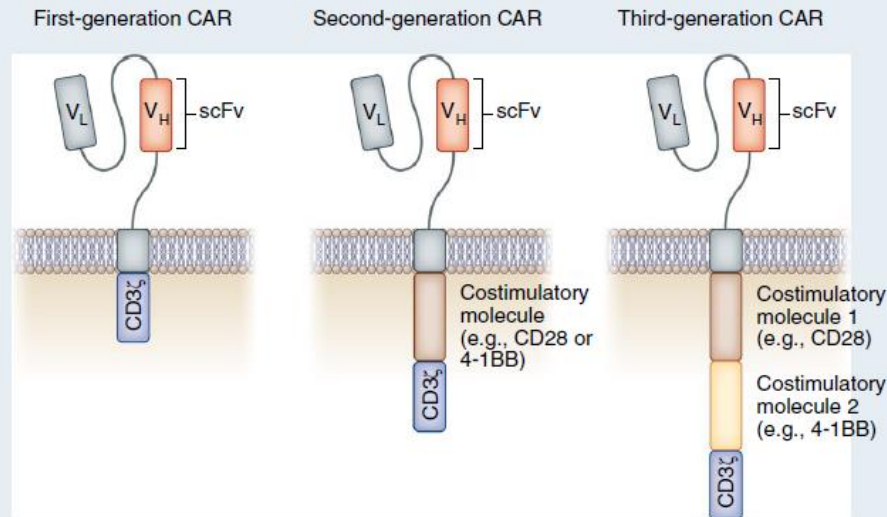
### Building a CAR

Chimeric antigen receptors are built by connecting several functional parts from different proteins, each with a specific job. A single-chain antibody variable fragment (Fv) recognizes a known cancer cell protein, targeting the tumor. These are generally engineered from monoclonal [antibodies](#) by using only the domains at the tip of the antibody, and connecting the two chains with a flexible linker. The antibody portion is connected to a transmembrane segment with another flexible linker. Inside the cell, one or more domains are taken from signaling proteins, which will activate the T cell once it finds a tumor cell.





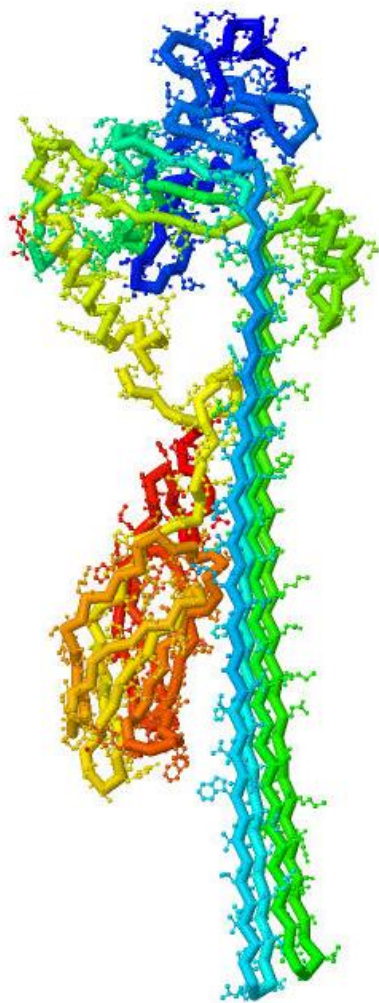
Structure of CAR T cells. CAR T cells are engineered by fusing the scFv of a monoclonal antibody (or another antigen-recognition domain) to a transmembrane domain and intracellular signaling domains capable of eliciting a T cell response.  $V_H$ , heavy chain;  $V_L$ , light chain; costim, costimulation; TM, transmembrane. Credit: Debbie Maizels/Springer Nature



CARs that contain only the CD3- $\zeta$  endodomain are known as first-generation CARs; those that contain one costimulatory domain (such as CD28 or 4-1BB) are known as second-generation CARs; and those that contain two or more costimulatory domains are known as third-generation CARs. Credit: Debbie Maizels/Springer Nature

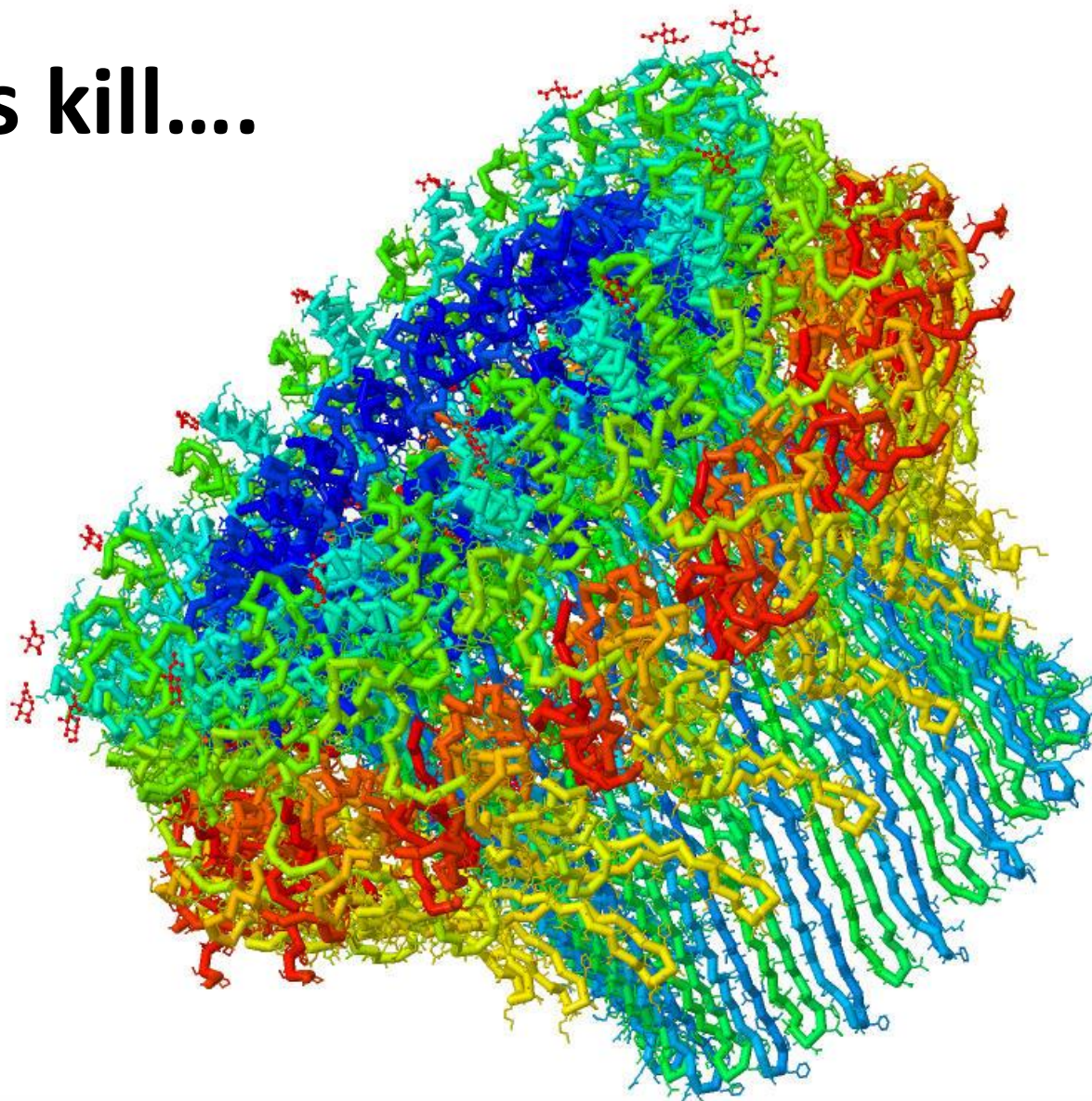
Perforin

# How T-cells kill....



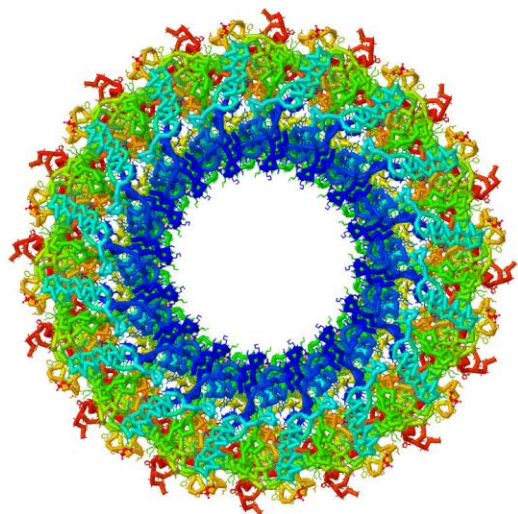
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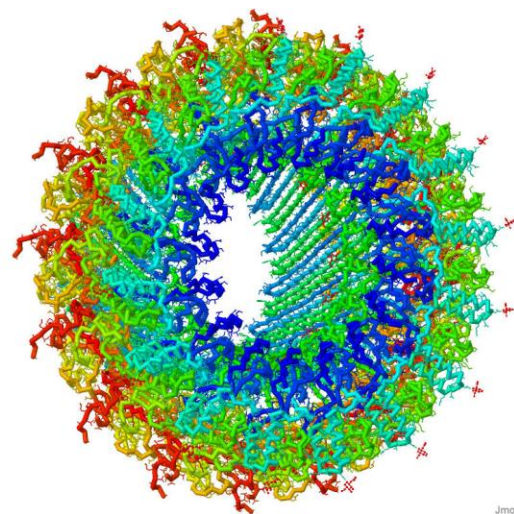


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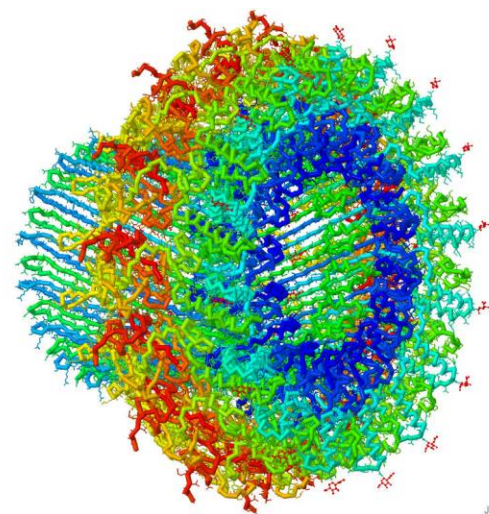
*models give meaning to words...*



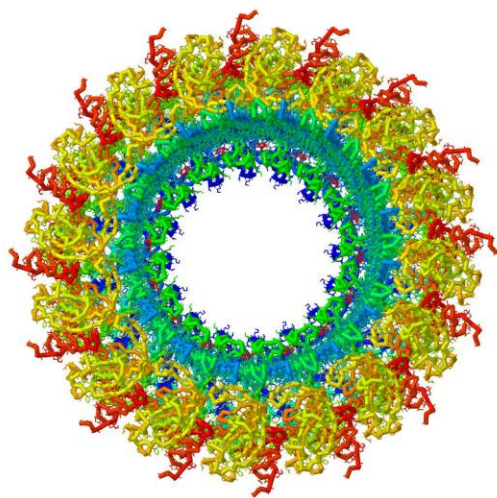
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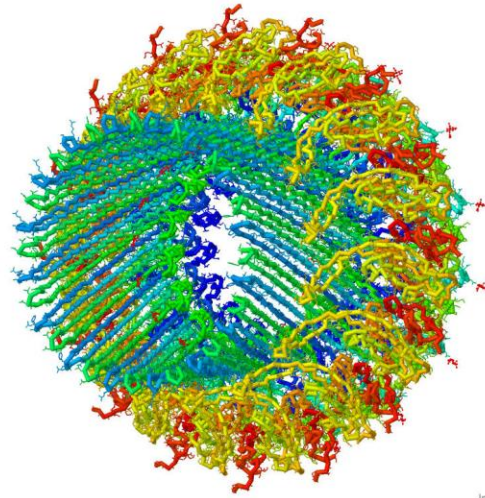
Jmol



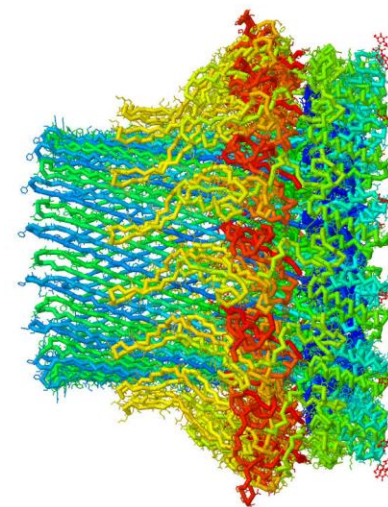
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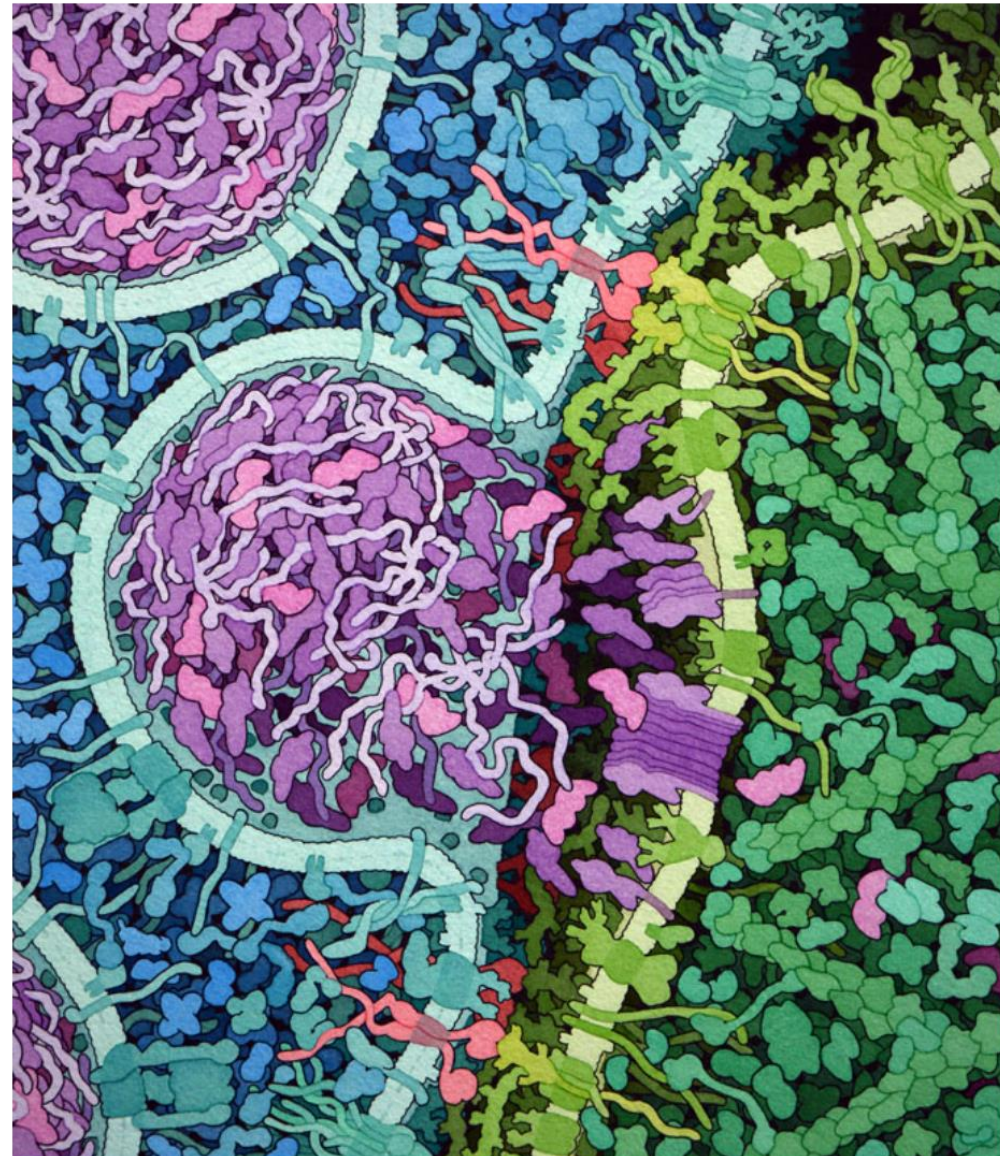


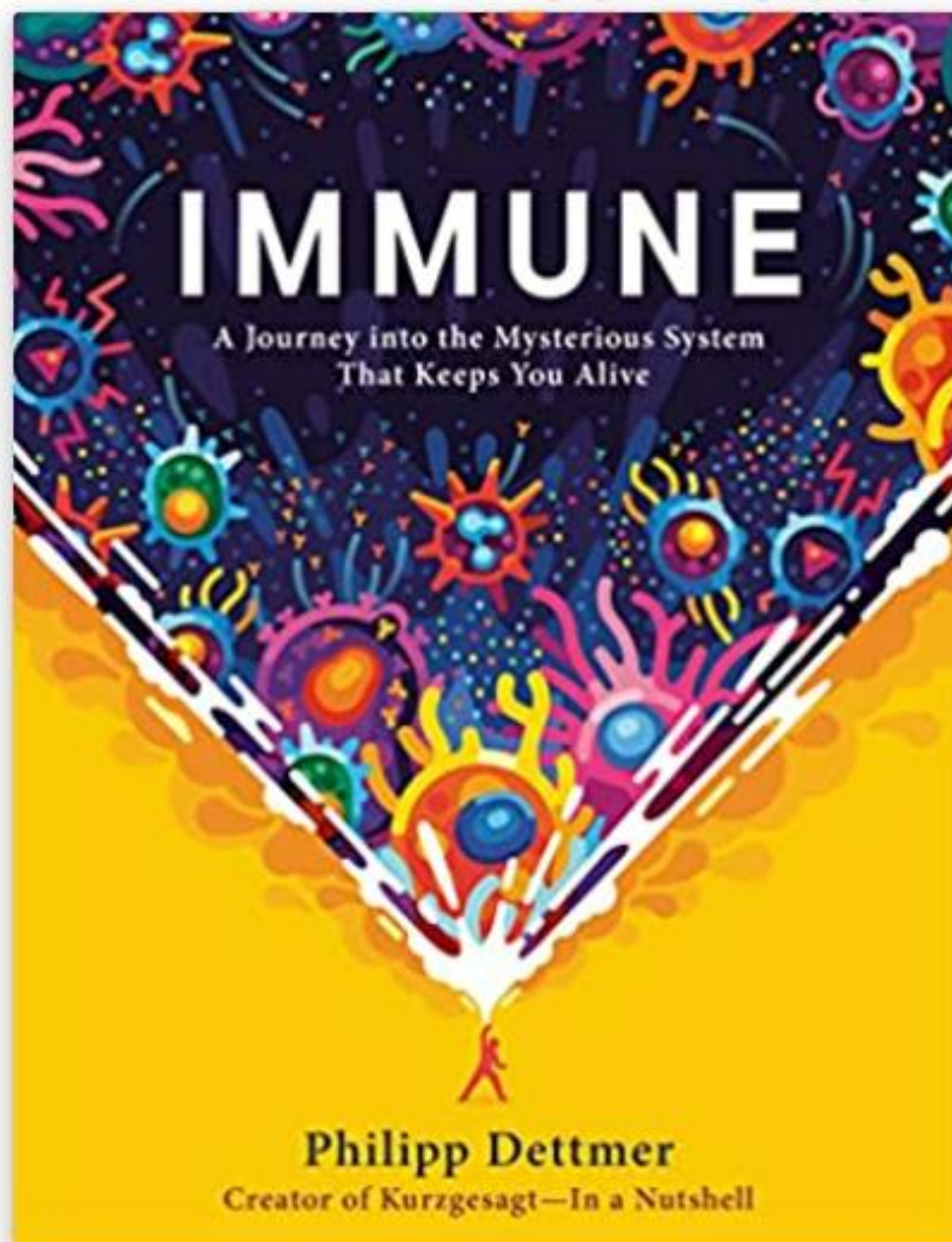
Jmol

## ***Models make it real....***

T-cells deploy perforin ... which spontaneously assembles into a tube that punches a hole in the cancer cell ... through which granzymes enter to trigger apoptosis of the cancer cell.

*Is science cool,... or what?*





### Milestones advisors

A 3D visualization of a cell. It features a large, textured blue sphere on the left, representing the nucleus, and a smaller, smooth orange sphere on the right, also representing a nucleus. Both spheres are connected by a narrow, elongated orange structure. Surrounding these spheres is a dense, intricate network of thin, blue, filamentous structures that radiate outwards, resembling a complex web or a network of fibers. The entire structure is set against a black background.

## Contents

|                                                                                                         |                                                                                                                                      |
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| <b>S2 Foreword</b>                                                                                      | organ-specific migration is identified                                                                                               |
| <b>S3 Thymus, a beginning and end 1961</b> The immunological function of the thymus gland is discovered | <b>S10 Identifying and cloning the T cell receptor 1984</b> The antigen receptor of the T cell is discovered                         |
| <b>S4 T cells as killers 1970</b> T cells are shown to kill cell targets                                | <b>S11 Surprising new TCR: the <math>\gamma\delta</math> T cells 1984</b> Identification of $\gamma\delta$ T cells                   |
| <b>S5 Discovery of T cell memory 1971</b> T cells contribute to immunological memory                    | <b>S12 T cell response to antigen is enabled by costimulation 1986</b> Discovery that T cells need costimulation for full activation |
| <b>S6 MHC restriction — why do</b>                                                                      |                                                                                                                                      |

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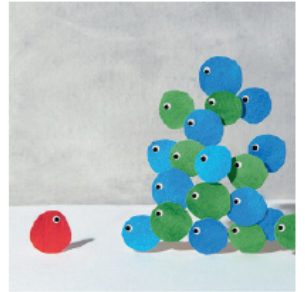
Alessandro Sette, La Jolla Institute  
for Immunology, USA

Brigitta Stockinger,  
The Francis Crick Institute, UK

E. John Wherry,  
University of Pennsylvania, USA

**Contributing journals**  
Communications Biology,  
Nature, Nature Communications,  
Nature Immunology,  
Nature Methods,

**Citing the Milestones**  
*Nature Milestones: T cells* includes Milestone articles written by our editors and an online collection of previously published material. To cite the full project, please use *Nature Milestones: T cells* <https://www.nature.com/collections/t-cells-milestones> (2022). Should you wish to cite any of the individual



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**Coming soon,... to a workshop near you.**

**Tomorrow's Science Today: *preparing for the next pandemic.***

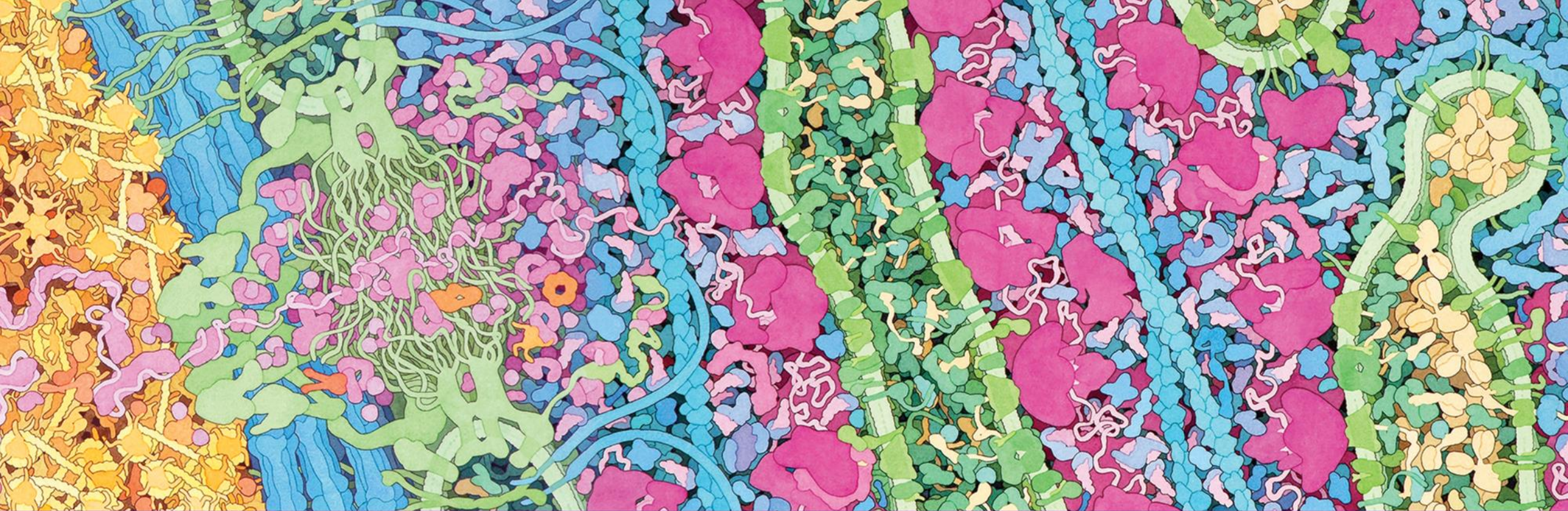
*An NIH SEPA project.*

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3d molecular  
designs

Transforming Science Learning

## RESEARCH ARTICLE SUMMARY

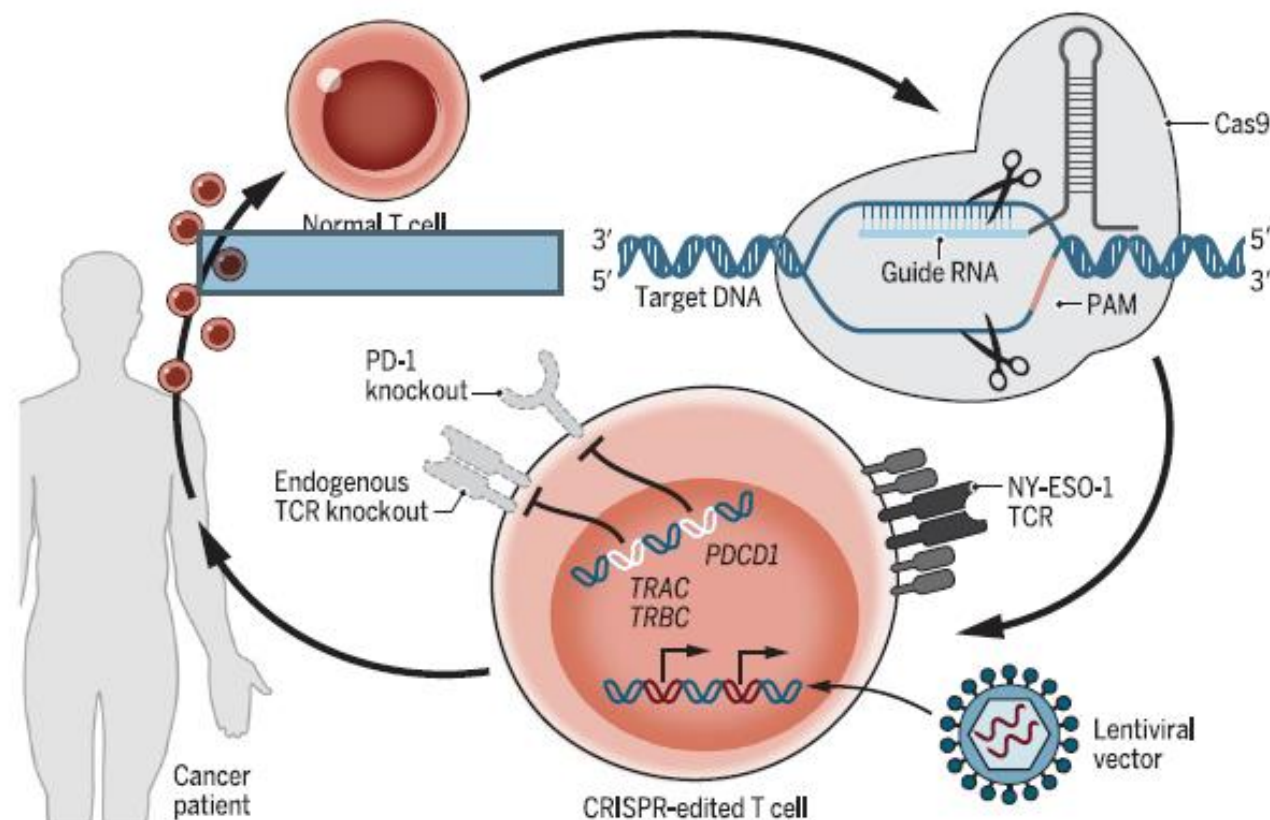
### CLINICAL TRIALS

# CRISPR-engineered T cells in patients with refractory cancer

Edward A. Stadtmauer<sup>\*†</sup>, Joseph A. Fraietta<sup>\*</sup>, Megan M. Davis, Adam D. Cohen, Kristy L. W. Eric Lancaster, Patricia A. Mangan, Irina Kulikovskaya, Minnal Gupta, Fang Chen, Lifeng Tia Vanessa E. Gonzalez, Jun Xu, In-young Jung, J. Joseph Melenhorst, Gabriela Plesa, Joanne Tina Matlawski, Amanda Cervini, Avery L. Gaymon, Stephanie Desjardins, Anne Lamontagn January Salas-McKee, Andrew Fesnak, Donald L. Siegel, Bruce L. Levine, Julie K. Jadowsky Regina M. Young, Anne Chew, Wei-Ting Hwang, Elizabeth O. Hexner, Beatriz M. Carreno, Christopher L. Nobles, Frederic D. Bushman, Kevin R. Parker, Yanyan Qi, Ansuman T. Satpat Howard Y. Chang, Yangbing Zhao, Simon F. Lacey<sup>\*</sup>, Carl H. June<sup>\*†</sup>

**INTRODUCTION:** Most cancers are recognized and attacked by the immune system but can progress owing to tumor-mediated immunosuppression and immune evasion mechanisms. The infusion of ex vivo engineered T cells, termed adoptive T cell therapy, can increase the natural antitumor immune response of the patient. Gene therapy to redirect immune specificity combined with genome editing has the potential to improve the efficacy and increase the safety of engineered T cells. CRISPR coupled with CRISPR-associated protein 9 (Cas9) endonuclease is a powerful gene-editing technology that potentially allows the ability to target multiple genes in T cells to improve cancer immunotherapy.

**RATIONALE:** Our first-in-human, phase 1 trial (clinicaltrials.gov; trial NCT0339 designed to test the safety and feasibility of multiplex CRISPR-Cas9 gene editing from patients with advanced, refractory cancer. A limitation of adoptively transferred T cells has been the induction of T cell exhaustion. We hypothesized that removing the endogenous T cell receptor (TCR) and the immune checkpoint molecule programmed cell death protein 1 (PD-1) would improve the function and persistence of engineered T cells. In addition, the removal of the endogenous TCR has the potential to improve safety and reduce toxicity that can be caused by auto-

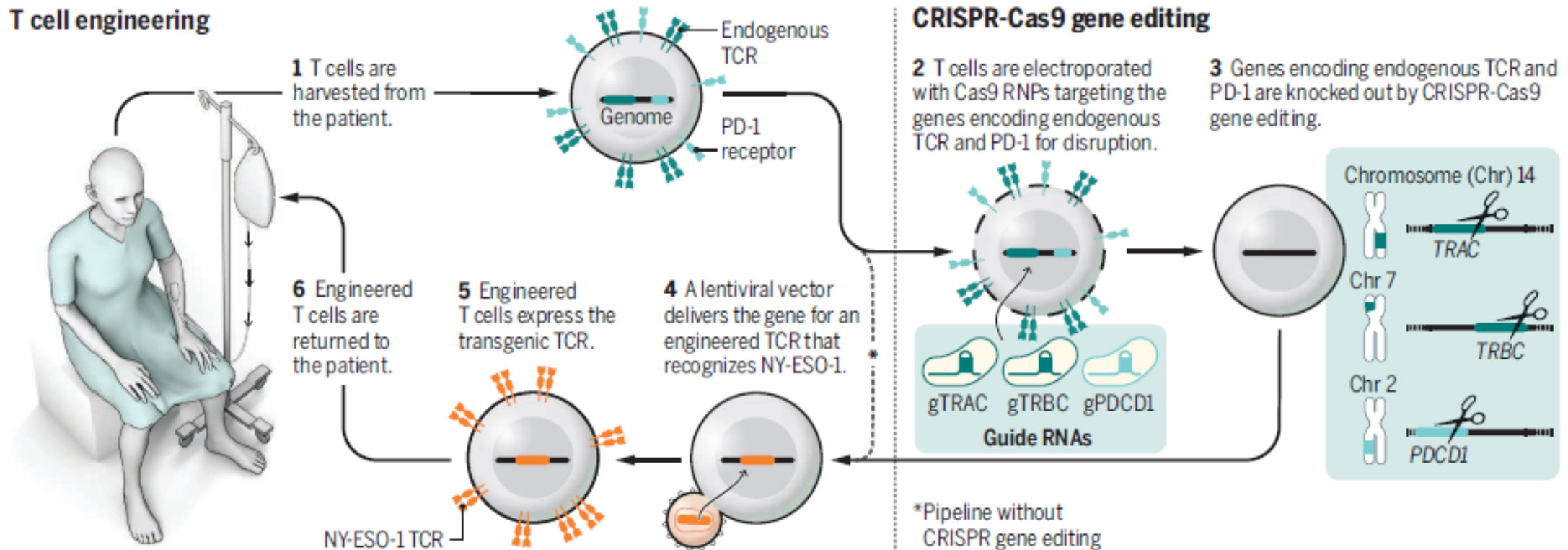


**CRISPR-Cas9 engineering of T cells in cancer patients.** T cells (center) were isolated from the blood of a patient with cancer. CRISPR-Cas9 ribonuclear protein complexes loaded with three sgRNAs were electroporated into the normal T cells, resulting in gene editing of the *TRAC*, *TRBC1*, *TRBC2*, and *PDCD1* (encoding PD-1) loci. The cells were then transduced with a lentiviral vector to express a TCR specific for the cancer-testis antigens NY-ESO-1 and LAGE-1 (right). The engineered T cells were then returned to the patient by intravenous infusion, and patients were monitored to determine safety and feasibility. PAM, protospacer adjacent motif.

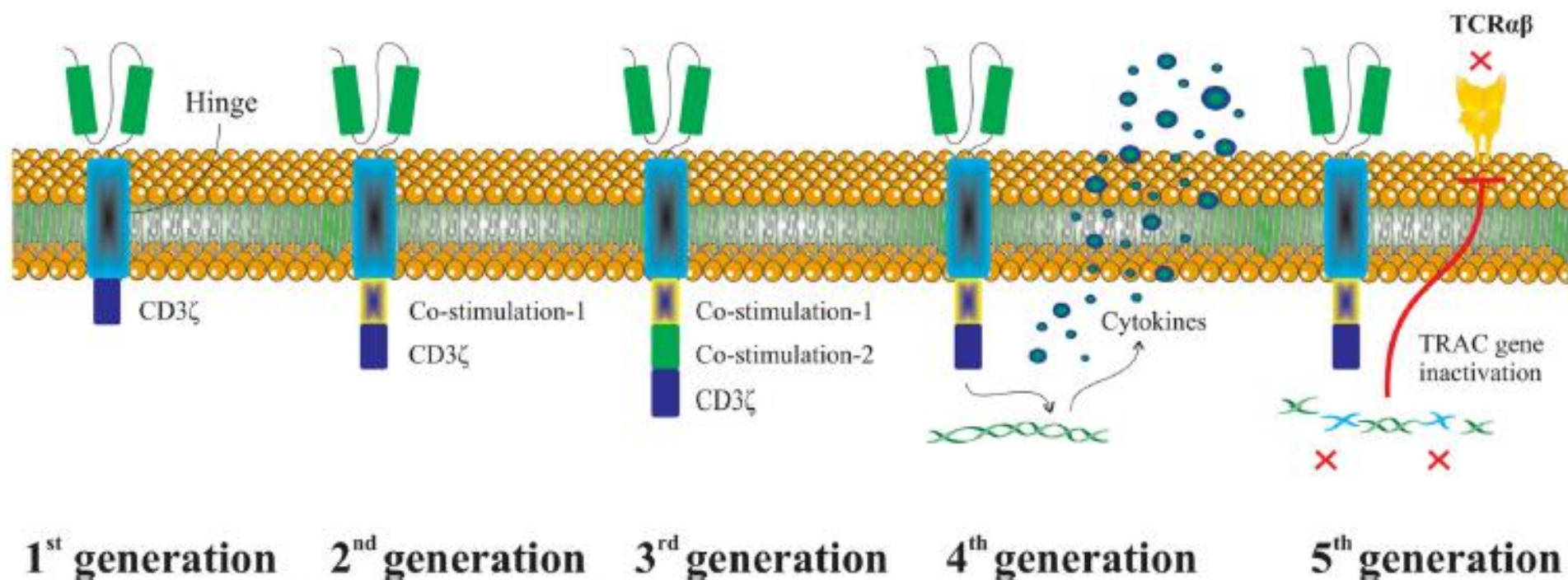
## Modifying engineered T cells with CRISPR-Cas9 gene editing

Engineered T cells with improved anticancer activity can be generated through the targeted disruption of immunomodulatory genes, such as programmed cell death protein 1 (*PDCD1*, which encodes PD-1), and T cell receptor (TCR) genes (*TRAC* and *TRBC*), using CRISPR-Cas9 delivered as preformed ribonucleoproteins (RNPs). These cells are then modified to express an engineered TCR that recognizes cancer-testis antigen 1 (NY-ESO-1) expressed by cancer cells.

### T cell engineering



# Generations of CAR T cells



**Fig. 1** The basic structure of CAR-T cell generations. The first generation of CARs contains only a CD3ζ as a well-documented intracellular signal domain. The second and third generations of CARs involve one or two costimulatory molecules in addition to CD3ζ, respectively. As well, the fourth generation of CAR-T cells strongly motivates the downstream transcription factor to prompt cytokine generation following interrelation between CAR and target antigen. Prominently, the genome edition technologies, such as CRISPR-Cas9, have been widely used to construct TRAC (TCR)-deficient CAR-T cells, establishing fifth generation of CAR-T cells